

Оригінальні дослідження

Original Researches

UDC 616.36-003.826

DOI: https://doi.org/10.22141/2224-0721.20.3.2024.1386

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Gut microbiota changes and novel markers associated with liver steatosis in obese patients

For citation: Mižnarodnij endokrinologičnij žurnal. 2024;20(3):179-184. doi: 10.22141/2224-0721.20.3.2024.1386

Abstract. Background. Liver steatosis is a common condition that can progress to steatohepatitis, fibrosis, and cirrhosis and increases the risk of death from cardiovascular and liver complications. Understanding the link between steatosis and non-alcoholic fatty liver disease, obesity, and gut microbiota is essential. Recent studies have revealed that gut microbiota plays a crucial role in developing this condition, highlighting the importance of microbiota control. The purpose of the study was to detect changes in gut microbiota and new markers associated with hepatic steatosis in obese patients. Materials and methods. The study involved 60 men aged 38 to 65, divided into two groups: 32 patients with hepatic steatosis (experimental group) and 28 with no steatosis (controls). As part of the study, the levels of the lipogram were determined, anthropometric measurements were made, a bioimpedance analysis of the body was performed, as well as liver ultrasound and shear wave elastography. The gut microbiota of all participants was also examined using sequencing technologies (material collected from stool samples). Results. In the experimental group, there are significantly more patients with overweight, dyslipidemia (hypercholesterolemia, triglyceridemia, high low-density lipoproteins, high atherogenicity coefficient, and low high-density lipoproteins). Also, patients with hepatic steatosis are more likely to have an excessive percentage of fat and an excessive amount of visceral fat, hepatomegaly due to the craniocaudal size of the liver, and increased liver stiffness. Regarding the intestinal microbiota, there is an increase in bacterial groups belonging to the Bacteroidetes. Our analysis showed that specific markers such as body mass index, blood lipid profile, body fat percentage, and liver ultrasound parameters are essential for diagnosing steatosis. Body mass index above 24.9 kg/m² and increased waist circumference were associated with steatosis. Bioimpedance analysis parameters, including body fat percentage and relative visceral fat level, were also crucial indicators. Dyslipidemia, with increased levels of total cholesterol, triglycerides, low-density lipoproteins, high atherogenicity coefficient, and lower high-density lipoproteins, was related to steatosis. The liver stiffness was significantly higher among patients with steatosis, indicating additional risk of liver fibrosis. Shear wave elastography can be a valuable tool for detecting liver steatosis. Conclusions. Patients with steatosis were characterized by signs of obesity (increased waist circumference, body mass index) and dyslipidemia, higher percentage of adipose tissue, relative amount of visceral fat, craniocaudal liver size, liver stiffness, and low levels of high-density lipoproteins. An increase in the gut microbiota of bacterial groups belonging to the Bacteroidetes has been observed.

Keywords: obesity; liver steatosis; intestinal microbiota

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Introduction

Liver steatosis represents a widespread condition that is a part of the general pathological process, which may lead to steatohepatitis under certain conditions and subsequently to liver fibrosis and even cirrhosis [1, 2]. It should be emphasized that patients with liver steatosis are at an increased risk of death from both cardiovascular diseases and liver-related complications [3].

The pathogenic role of steatosis in liver damage is recognized regardless of its origin. However, researchers are particularly interested in detecting steatosis in the context of non-alcoholic fatty liver disease due to its widespread occurrence and close association with metabolic disorders [4]. This is especially crucial as steatosis is highly prevalent among individuals with obesity, diabetes, and dyslipidemia [5]. Additionally, it is known that fatty liver degeneration can occur long before the development of metabolic syndrome. In contrast, liver steatosis may develop even with a slight increase in body weight, as in this case, redistribution of lipid content in tissues and metabolic disorders can occur [6].

Experimental studies have revealed that the bacteria living in our intestines have an equally important role in the development of metabolic dysfunction-associated fatty liver disease (MAFLD) [7]. Lifestyle changes and increased physical activity can alter the state of our liver through their influence on the gut microbiota composition. It is worth noting that the detected changes in the composition of the gut microbiota in patients with MAFLD can be managed by the use of particular drugs, such as probiotics, prebiotics, or synbiotics, allowing to improve the condition of patients with MAFLD significantly [8].

The timely detection and quantification of both the state of the liver and the gut microbiota have become crucial objectives for hepatology, endocrinology, and other areas of the healthcare system overall, given the high prevalence of steatosis.

The purpose of the study was to identify gut microbiota changes and novel markers associated with liver steatosis in obese patients.

Materials and methods

Sixty men aged 38 to 65 years (average of 48.6 ± 10.8 years) were involved in the study. Non-inclusion criteria were the presence of alcoholic, drug-induced, viral, autoimmune liver damage, and storage diseases.

All patients were examined for the presence of fatty hepatosis according to the European Association for the Study of the Liver — European Association for the Study of Diabetes — European Association for the Study of Obesity Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease [1] using liver ultrasound in the absence of apparent reasons for the secondary build-up of fat in the liver (alcohol abuse, use of hepatotoxic drugs, viral hepatitis infection, autoimmune and hereditary diseases).

Exclusion of alcoholic liver damage was performed based on reports of daily alcohol consumption in a dose of at least 50.0 g of pure ethanol for 25 years and screening for alcohol abuse (CAGE questionnaire — Cut down, Annoyed, Guilty, Eye-opener). Hepatitis B virus etiology was identified using serological markers of hepatitis B virus (HBsAg determina-

tion), hepatitis C virus — serological markers of hepatitis C virus (anti-HCV), as well as information regarding acute viral hepatitis in the previous medical records. Autoimmune reactions were evaluated based on the serological markers of autoimmune liver damage.

Two groups were formed depending on the presence of steatosis: experimental (EG) - 32 people with ultrasound-confirmed steatosis, and control (CG) - 28 patients without steatosis.

Biochemical blood serum parameters were studied using an automatic analyzer, Beckman Coulter AU680 (USA), which detected the levels of total cholesterol, triglycerides, high-density lipoproteins (HDL), and low-density lipoproteins (LDL). The atherogenicity coefficient (AC) greater than 3 was considered to be elevated.

Bioimpedance analysis was conducted using Tanita BC-587 body composition analyzer scales (Tanita, Japan) to determine patients' body weight, body mass index (BMI) (kg/m²), bone mass (kg), skeletal muscle mass (kg), water content (%), body fat percentage (%), visceral (internal) fat level (units), basic metabolism (kcal), biological age (calculated based on the body's metabolic rate) (years). We used the following indicators for the study: body fat percentage and visceral fat level. The upper limits of the normal reference range accepted during bioimpedance analysis in the studied age group were as follows: for the body fat — 20 % and less, visceral (internal) fat levels — 12 and less, whereas BMI — 24.9 kg/m² and less. Patients' waist circumference (WC) was also measured as the primary criterion for metabolic disorders. WC exceeding 102 cm was considered elevated.

A liver ultrasound examination was conducted using a Toshiba (Canon) Aplio 500 Platinum device with a 2–5 MHz convex probe under standard conditions: in the morning, on an empty stomach, in a horizontal position lying on the back. All patients underwent standard ultrasound to identify functional and structural changes in the internal organs; liver dimensions (mm), outlines, structure, and echogenicity were assessed.

Shear wave elastography was performed on a Toshiba (Canon) Aplio 500 Platinum device in 2D diagnostic mode using a CA1-7A probe. The measurements were taken in real-time at a depth of 20–60 mm from the capsule to determine the average indicators that characterize the stiffness of the liver parenchyma (kPa). Threshold values were used to assess the stage of fibrosis: F0 — no fibrosis (up to 5.8 kPa); F1 — mild fibrosis (5.9–7.2 kPa); F2 — moderate fibrosis (7.3–9.5 kPa); F3 — advanced fibrosis (9.6–12.5 kPa); F4 — liver cirrhosis (> 12.5 kPa) according to the METAVIR scale.

The gut microbiota was evaluated by determining the fecal bacterial composition using a polymerase chain reaction-based sequencing method in a certified laboratory DIA-GEN, which provided results in the form of metagenomic analysis. The most used method for this purpose is 16S rRNA gene sequencing. This technique targets a specific region of the bacterial 16S rRNA gene, allowing for the identification and classification of bacterial species present in the sample collected from the stool.

The results were processed in Microsoft Excel using descriptive statistics, Student's t-test, and z-criterion to compare two variables. The statistical reliability of the markers



was evaluated using the SPSS software based on contingency tables, and indicators of diagnostic value were calculated. The correlation between steatosis and the indicator studied was considered confirmed by a module if the association coefficient exceeded 0.5 (or 0.3 for the contingency coefficient). The results were presented as M \pm m, where M is the arithmetic mean, m is the mean square deviation; n is the number of patients examined in the group. The difference was considered statistically significant if p < 0.05.

Results

Patients of the EG had significantly higher levels of total cholesterol compared to the CG (6.5 \pm 1.4 mmol/l vs. 4.0 \pm 0.6 mmol/l). In addition, significantly higher triglycerides (2.3 \pm 0.8 mmol/l vs. 1.1 \pm 0.1 mmol/l), LDL, and AC were observed in the group with liver steatosis. The mean value of HDL was not significantly different in both groups of patients. The results are shown in Table 1.

According to the information presented in Table 2, WC in patients with hepatic steatosis was significantly greater than in the CG (109.0 \pm 12.0 cm vs. 97.6 \pm 9.8 cm). BMI was also significantly higher in the group with steatosis compared to the CG patients (28.0 \pm 2.7 kg/m² vs. 21.6 \pm 1.7 kg/m²), as was the body fat percentage and the relative visceral fat level (32.3 \pm 6.1 % vs. 20.9 \pm 4.2 % and 9.4 \pm 3.3 units vs. 4.0 \pm 1.5 units, respectively).

The liver ultrasound results (Table 3) indicate a significantly greater craniocaudal (right) liver lobe length in patients with steatosis compared to the CG (160.5 \pm 11.0 mm vs. 130.4 \pm 13.4 mm) and a significantly higher index of liver stiffness in the EG compared to the CG (5.5 \pm 0.6 kPa vs. 3.8 \pm 1.3 kPa). The left liver dimension was not significantly different between the groups of patients (60.3 \pm 5.6 mm vs. 57.1 \pm 6.5 mm).

We obtained the following results for each group after comparing the number of patients with indicators exceeding the reference range. The number of patients with increased WC was higher in the EG, but it was not significant (19/32 vs. 11/28). However, a significantly more patients in the

Table 1. Blood lipid profile

Parameter	EG, n = 32	CG, n = 28
Total cholesterol, mmol/L	6.5 ± 1.4*	4.0 ± 0.6
Triglycerides, mmol/L	2.3 ± 0.8*	1.1 ± 0.1
LDL, mmol/L	4.6 ± 1.2*	2.8 ± 0.3
HDL, mmol/L	1.1 ± 0.2	1.2 ± 0.5
AC, units	5.2 ± 1.8*	2.7 ± 1.0

Note: here and in Tables 2, 3: * — p < 0.05 compared to CG values.

Table 2. Results of anthropometry and bioimpedance analysis

Parameter	EG, n = 32	CG, n = 28
WC, cm	109.0 ± 12.0*	97.6 ± 9.8
BMI, kg/m ²	28.0 ± 2.7*	21.6 ± 1.7
Body fat percentage, %	32.3 ± 6.1*	20.9 ± 4.2
Visceral fat level, units	9.4 ± 3.3*	4.0 ± 1.5

Table 3. Liver ultrasound and shear wave elastography

Parameter	EG, n = 32	CG, n = 28
Craniocaudal (right) lobe length, mm	160.5 ± 11.0*	130.4 ± 13.4
Left liver size, mm	60.3 ± 5.6	57.1 ± 6.5
Liver stiffness index, E, kPa	5.5 ± 0.6*	3.8 ± 1.3

EG had high BMI (28/32 vs. 1/28), high levels of total cholesterol (27/32 vs. 1/28), triglycerides (24/32 vs. 6/28), high AC (30/32 vs. 8/28), high body fat percentage (31/32 vs. 8/28). There were significantly more patients with an insufficient HDL level in the EG compared to CG (29/32 vs. 15/28). Higher levels of LDL compared to the reference range (14/32), higher than normal relative visceral fat level

Table 4. Analysis of the diagnostic value of additional screening markers of the liver steatosis risk

Parameter	Sensitivity, %	Specificity, %	Accuracy, %	Association/contingency coefficient
WC	60.0	60.0	60.0	0.38
ВМІ	87.5	95.0	90.0	0.98*
Total cholesterol	82.5	95.0	86.7	0.98*
Triglycerides	75.0	80.0	76.7	0.85*
LDL	42.5	100.0	61.7	0.44*
HDL	90.0	45.0	75.0	0.76*
AC	92.5	70.0	85.0	0.93*
Body fat percentage	97.5	70.0	88.3	0.98*
Visceral fat level	27.5	100.0	51.7	0.33*
Craniocaudal (right) liver length	72.5	100.0	81.7	0.68*
Left liver size	_	_	_	-
Liver stiffness	32.5	90.0	51.7	0.63*

Note: * — statistically confirmed correlation between the marker and the risk of liver steatosis.



(9/32), and increased craniocaudal size of the liver were only observed among patients of the EG (23/32). All the above indicators were within the reference range in controls. The left liver size was not increased in patients of both groups.

The following research phase is statistical analysis, which calculates the chances of accurately diagnosing liver steatosis using individual indicators — markers. Indicators of diagnostic value, such as sensitivity, specificity, and accuracy, were employed to determine the reliability of the diagnosis. Based on them, the odds ratio was defined — a number that shows how much the absence or presence of a particular result is associated with the presence or absence of a specific disease in a statistical group, and the association (or contingency) coefficient, which characterizes how close is the stochastic association between qualitative signs — alternative random variables. The sensitivity, specificity, accuracy, and association (or contingency) coefficient are shown in Table 4.

Based on the obtained data, WC, although it is a marker of metabolic disorder, did not indicate the presence of liver steatosis. However, all other assessed indicators can be considered risk markers of liver steatosis. Namely, BMI with a sensitivity of 87.5 %, specificity of 95.0 %, and accuracy of 90.0 % indicates the presence of liver steatosis. Blood lipid indicators are also of high importance: total cholesterol with a sensitivity of 82.5 %, specificity of 95.0 %, and accuracy of 86.7 % indicates the presence of steatosis, as well as triglycerides with a sensitivity of 75.0 %, specificity of 80.0 %, accuracy of 76.7 %; LDL with a sensitivity of 42.5 %, specificity of 100.0 %, accuracy of 61.7 %; HDL with a sensitivity of 90.0 %, specificity of 45.0 %, accuracy of 75.0 %; AC with a sensitivity of 92.5 %, specificity of 70.0 %, and accuracy of 85.0 %. The indicators of bioimpedance analysis, specifically the percentage of body fat tissue and the relative visceral fat level, are equally important. Excess visceral fat level with a sensitivity of 97.5 %, specificity of 70.0 %, and accuracy of 88.3 %, and relative visceral fat level with a sensitivity of 27.5 %, specificity of 100.0 % and accuracy of 51.7 % indicate liver steatosis. Regarding the liver ultrasound results, the craniocaudal size of the liver indicated hepatic steatosis with a sensitivity of 72.5 %, specificity of 100.0 %, and accuracy of 81.7 %. In contrast, the liver stiffness index had a sensitivity of 32.5 %, specificity of 90.0 %, and accuracy of 51.7 %.

Gut microbiota in obese patients also demonstrated a shift in the distribution of bacterial composition compared to the CG.

Patients of the EG had a higher total bacterial mass, which exceeded the data of controls and conditionally established levels of the reference range ($< 10^{12}$ CFU/cm³): 2.51 ± 0.21 CFU/cm³ vs. 1.72 ± 0.19 CFU/cm³ (p < 0.05). A more detailed review of the quantitative proportional distribution of the total bacterial mass according to the polymerase chain reaction in the EG showed that more than half of the bacteria belonged to the *Bacteroidetes* type (51.49 ± 4.16 %). In comparison, all the other types of bacteria in the smaller half of the total bacterial mass were distributed in the following proportional composition: *Firmicutes* — 32.42 ± 2.14 %, *Actinobacteria* — 6.12 ± 1.94 %,

other groups -9.61 ± 2.18 %. Thus, a shift towards dysbiosis in the Firmicutes/Bacteroidetes ratio (0.62 \pm 0.21) was also observed (relative norm 1.0-5.0). No significant changes were noted during the examination of the CG patients in the distribution of gut microbiota; accordingly, the Firmicutes/Bacteroidetes ratio did not reflect significant deviations and was 1.12 ± 0.44 , which corresponds to the reference range. Bacteroides fragilis/Faecalibacterium prausnitzii ratio (reference range < 100.0) in patients of the EG significantly exceeded the established physiological reference range due to an increase in the Bacteroides fragilis group (1121.02 \pm 847.13), which in some patients exceeded the physiological reference range by more than 15 times. It was within the normal range only in 21.17 % of patients, while in the CG, the above indicator did not exceed the reference range in more than 62 %.

Discussion

In our study, we focused on analyzing the link between the presence of liver steatosis and several indicators that directly or indirectly indicate the presence of a metabolic disorder. Our choice was not accidental, as it is known that non-alcoholic fatty liver disease and liver steatosis, which is one of its components, are pathogenetically closely linked [9, 10]. This topic is of paramount importance since there is an ongoing discussion among scientists worldwide regarding redefining the term "non-alcoholic fatty liver disease" to "metabolically associated liver disease", thereby transferring the disease from the rank of diseases of exclusion to a nosology with clearly defined diagnostic criteria [11].

It's not just the scientific aspect that makes this topic relevant. As of 2015, around 604 million people across the globe are suffering from obesity [12]; the prevalence of type 2 diabetes and hypertension also contributes to the development of liver steatosis, with a risk of transforming into steatohepatitis and later into liver cirrhosis and, in some cases, hepatocellular carcinoma [13]. All of this underscores the urge for early detection of liver steatosis.

Our statistical analysis has confirmed the importance of specific markers such as BMI greater than 24.9 kg/m², blood lipid indicators, body fat percentage, and relative visceral fat level, along with specific liver ultrasound parameters (craniocaudal liver size and liver stiffness index).

In addition to the significant practical value, the data we obtained provide a specific understanding of the "risk group" — patients most likely to develop steatosis. Our results emphasize that despite increased WC is an important marker of metabolic disorder, it did not become the factor that determined the presence of hepatic steatosis. In contrast to WC, BMI > 24.9 kg/m² was reliably associated with steatosis. Bioimpedance analysis parameters (body fat percentage and relative visceral fat level) were crucial. The above data highlights the importance of conducting a comprehensive examination rather than focusing on the visible deposition of fatty tissue in certain areas.

Blood lipid indicators, such as increased levels of total cholesterol, triglycerides, LDL, high AC, and lower HDL, are directly pathogenetically related to steatosis. This indicates that in dyslipidemia, the patient should undergo a comprehensive examination with screening for liver steatosis.



Regarding the gut microbiota, the predominance of *Bacteroidetes* over all groups of bacteria is noteworthy. An interesting feature was the predominance of this type of bacteria in patients with hepatic steatosis compared to other groups and across the total population combined. The identified changes raise many concerns, namely, whether such a redistribution of the gut microbiota composition is a consequence of the deterioration of the metabolic status of the liver and obesity, or possibly the opposite, an extreme increase in bacteria of the *Bacteroidetes* type is capable of triggering liver steatosis and obesity [14, 15].

The significance of liver stiffness in liver ultrasound cannot be overstated, as it was significantly higher among patients with steatosis. Additionally, exceeding the threshold values was often observed among participants with steatosis compared to those without it, which indicates additional risks of liver fibrosis in such patients [16–18]. The routine performance of shear wave elastography can serve as a valuable tool in determining liver steatosis.

Conclusions

Patients diagnosed with hepatic steatosis had significantly higher WC and BMI, as well as higher levels of total cholesterol, triglycerides, LDL, and AC, higher body fat percentage and relative visceral fat level, significantly larger craniocaudal (right) liver size, and higher liver stiffness when compared to those without this condition. However, the mean value of HDL did not differ significantly between two groups.

In the group with hepatic steatosis, there were significantly more patients with increased BMI, high levels of total cholesterol, triglycerides, LDL, AC, and low HDL. They also had an increase in the following parameters: body fat percentage and relative visceral fat level, craniocaudal liver size, and liver stiffness. Furthermore, the growth of *Bacteroidetes* groups was observed in the gut microbiota.

Markers associated with the presence of hepatic steatosis include high BMI, total cholesterol, triglycerides, LDL, AC, bioimpedance analysis indicators, namely the body fat percentage and the relative visceral fat level, certain parameters of the liver ultrasound, such as craniocaudal liver size and liver stiffness index, and low HDL levels.

References

- 1. European Association for the Study of the Liver (EASL); European Association for the Study of Diabetes (EASD); European Association for the Study of Obesity (EASO). EASL-EASD-EASO Clinical Practice Guidelines for the management of non-alcoholic fatty liver disease. J Hepatol. 2016 Jun;64(6):1388-402. doi:10.1016/j. jhep.2015.11.004.
- 2. Pouwels S, Sakran N, Graham Y, et al. Non-alcoholic fatty liver disease (NAFLD): a review of pathophysiology, clinical management and effects of weight loss. BMC Endocr Disord. 2022 Mar 14;22(1):63. doi:10.1186/s12902-022-00980-1.
- 3. Lee KC, Wu PS, Lin HC. Pathogenesis and treatment of non-alcoholic steatohepatitis and its fibrosis. Clin Mol Hepatol. 2023 Jan;29(1):77-98. doi:10.3350/cmh.2022.0237.
 - 4. Kasper P, Martin A, Lang S,et al. NAFLD and cardiovascular

- diseases: a clinical review. Clin Res Cardiol. 2021 Jul;110(7):921-937. doi:10.1007/s00392-020-01709-7.
- 5. Romero-Gómez M. Non-alcoholic steatohepatitis. Med Clin (Barc). 2022 Oct 28;159(8):388-395. English, Spanish. doi:10.1016/j. medcli.2022.06.017.
- 6. Solomentseva TA. New criteria for metabolic dysfunction-associated fatty liver disease: advantage or question? Review. Mod Gastroenterol. 2023 Aug 31;(4):84-90. doi:10.30978/mg-2023-4-84.
- 7. Safari Z, Gérard P. The links between the gut microbiome and non-alcoholic fatty liver disease (NAFLD). Cell Mol Life Sci. 2019 Apr;76(8):1541-1558. doi:10.1007/s00018-019-03011-w.
- 8. Komarytsia OY, Radchenko OM, Moskva KA, Borovets MO. Changes in gut microbiota in patients with metabolic-associated fatty liver disease. Mižnarodnij endokrinologičnij žurnal. 2023 Oct 31;19(6):419-23. doi:10.22141/2224-0721.19.6.2023.1309. (in Ukrainian).
- 9. Barlow GM, Mathur R. Type 2 Diabetes and the Microbiome. J Endocr Soc. 2022 Nov 30;7(2):bvac184. doi:10.1210/jendso/bvac184.
- 10. Castaner O, Goday A, Park YM, et al. The Gut Microbiome Profile in Obesity: A Systematic Review. Int J Endocrinol. 2018 Mar 22;2018:4095789. doi:10.1155/2018/4095789.
- 11. van Son J, Koekkoek LL, La Fleur SE, Serlie MJ, Nieuwdorp M. The Role of the Gut Microbiota in the Gut-Brain Axis in Obesity: Mechanisms and Future Implications. Int J Mol Sci. 2021 Mar 15;22(6):2993. doi:10.3390/ijms22062993.
- 12. Muscogiuri G, Cantone E, Cassarano S, et al; on behalf of the Obesity Programs of nutrition, Education, Research and Assessment (OPERA) group. Gut microbiota: a new path to treat obesity. Int J Obes Suppl. 2019 Apr;9(1):10-19. doi:10.1038/s41367-019-0011-7.
- 13. Marzullo P, Bettini S, Menafra D, et al; Obesity Programs of nutrition, Education, Research and Assessment (OPERA) group. Spot-light on microbiota in obesity and cancer. Int J Obes (Lond). 2021 Nov;45(11):2291-2299. doi:10.1038/s41366-021-00866-7.
- 14. Ballini A, Scacco S, Boccellino M, Santacroce L, Arrigoni R. Microbiota and Obesity: Where Are We Now? Biology (Basel). 2020 Nov 25;9(12):415. doi:10.3390/biology9120415.
- 15. Liu BN, Liu XT, Liang ZH, Wang JH. Gut microbiota in obesity. World J Gastroenterol. 2021 Jul 7;27(25):3837-3850. doi:10.3748/wjg.v27.i25.3837.
- 16. Tokarek J, Gadzinowska J, Młynarska E, Franczyk B, Rysz J. What Is the Role of Gut Microbiota in Obesity Prevalence? A Few Words about Gut Microbiota and Its Association with Obesity and Related Diseases. Microorganisms. 2021 Dec 27;10(1):52. doi:10.3390/microorganisms10010052.
- 17. Tkach SM, Pankiv VI, Krushinska ZH. Features of type 2 diabetes combined with metabolic dysfunction-associated fatty liver disease under conditions of chronic stress. Mižnarodnij endokrinologičnij žurnal. 2024:20(1):18-24. doi:10.22141/2224-0721.20.1.2024.1353. (in Ukrainian).
- 18. Miloslavsky DK, Koval SM. Prospects for probiotics use as the gut microbiota modulators in obesity (literature review). Mižnarodnij endokrinologičnij žurnal. 2022;18(6):358-364. doi:10.22141/2224-0721.18.6.2022.1207. (in Ukrainian).

Received 19.02.2024 Revised 18.04.2024 Accepted 30.04.2024 ■



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Conflicts of interests. Authors declare the absence of any conflicts of interests and own financial interest that might be construed to influence the results or interpretation of the manuscript.

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Зміни мікробіоти кишечника та нові маркери, пов'язані зі стеатозом печінки, у пацієнтів з ожирінням

Резюме. *Актуальність*. Стеатоз печінки є поширеним ураженням, яке може прогресувати до стеатогепатиту, фіброзу та цирозу, і збільшує ризик смерті від серцево-судинних та печінкових ускладнень. Важливим є зв'язок стеатозу з неалкогольною жировою хворобою печінки, ожирінням та кишковою мікробіотою. У недавніх дослідженнях з'ясували, що кишкова мікробіота відіграє важливу роль у розвитку цього захворювання, тому контроль мікробіоти є актуальним завданням. Мета: виявити зміни кишкової мікробіоти та нові маркери, пов'язані зі стеатозом печінки, у пацієнтів з ожирінням. Матеріали та методи. У дослідження було залучено 60 чоловіків віком від 38 до 65 років, яких розділили на дві групи: 32 пацієнти зі стеатозом (експериментальна група) та 28 осіб без стеатозу (контрольна група). У рамках дослідження визначено рівні ліпідограми, здійснено антропометричні вимірювання, проведено біоімпедансний аналіз тіла, ультразвукове дослідження та зсувнохвильову еластографію печінки. Мікробіота кишечника всіх учасників також була досліджена за допомогою технологій секвенування (матеріал, зібраний зі зразків калу). Результати. В експериментальній групі було значно більше осіб із надмірною масою тіла, дисліпідемією (гіперхолестеринемія, тригліцеридемія, високі рівень ліпопротеїнів низької щільності й коефіцієнт атерогенності, низький уміст ліпопротеїнів високої щільності). Також у пацієнтів зі стеатозом печінки частіше спостерігають надмірні відсоток жиру і кількість віс-

церального жиру, гепатомегалію внаслідок краніокаудального розміру печінки, підвищену жорсткість печінки. Що стосується кишкової мікробіоти, то зареєстровано збільшення груп бактерій типу Bacteroidetes. Такі специфічні маркери, як індекс маси тіла, показники ліпідного профілю крові, відсоток жиру в організмі та ультразвукові параметри печінки, важливі для діагностики стеатозу. Індекс маси тіла понад 24,9 кг/м² і збільшена окружність талії пов'язані зі стеатозом. Параметри біоімпедансного аналізу, як-от відсоток жиру в організмі та відносний рівень вісцерального жиру, також ε вирішальними показниками. Дисліпідемія з підвищеними рівнями загального холестерину, тригліцеридів, ліпопротеїнів низької щільності і нижчим рівнем ліпопротеїнів високої щільності пов'язана зі стеатозом. Індекс жорсткості печінки вірогідно вищий серед пацієнтів зі стеатозом, що вказує на додаткові ризики фіброзу печінки. Зсувнохвильова еластографія може бути корисним інструментом при виявленні стеатозу печінки. Висновки. Для пацієнтів зі стеатозом характерними були ознаки ожиріння та дисліпідемія, підвіщені кількість жирової тканини, вісцерального жиру, краніокаудальний розмір печінки, показник жорсткості печінки й низькі рівні ліпопротеїдів високої щільності. Серед мікробіоти кишечника спостерігалося збільшення бактеріальних груп, що належать до типу Bacteroidetes.

Ключові слова: ожиріння; стеатоз печінки; мікробіота кишечника

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